

Interaction of Scots Pine Defensin with Model Membrane by Coarse-Grained Molecular Dynamics

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Abstract

© 2017, Springer Science+Business Media New York. Plant defensins are a part of the innate immune system of plants that acts against a broad range of pathogens. Many plant defensins, including pine defensins, show strong antifungal activity that is associated with their ability to penetrate into the fungal cell membrane. However, the exact molecular mechanism of their action remains poorly defined. To obtain insight into the mechanism of protein-membrane interaction, we applied a coarse-grained molecular dynamics simulation to study the interaction of pine defensin with two model membranes: the first consisted of zwitterion-neutral POPC molecules and the second was composed of combined anionic POPG and POPC. The simulations show that defensin does not form stable complexes with the neutral membrane but does interact with the combined POPG/POPC membrane. In the latter case, defensin attaches to the membrane surface by interacting with lipid polar heads without deep penetration into the hydrophobic tail zone. Electrostatic interactions are a driving force of the complex formation, which determines the orientation of the protein relative to the bilayer surface. Two favorable orientations of defensin are detected where the defensin molecule orients either perpendicular or parallel to the membrane plane. Being positively charged, pine defensin induces changes in the lipid distribution along the membrane, resulting in the formation of zones with different electrostatic potentials that can cause deformation or distortion of the membrane. Pine defensin is a representative of plant defensins, and hence the results of this study can be applied to other members of the family.

<http://dx.doi.org/10.1007/s00232-017-9950-5>

Keywords

Coarse-grained molecular dynamics, Defensin, Lipid-protein interactions, Membrane

References

- [1] Aerts AM, François IE, Bammens L, Cammue B, Smets B, Winderickx J, Accardo S, de Vos DE, Thevissen K (2006) Level of M(IP)2 C sphingolipid affects plant defensin sensitivity, oxidative stress resistance and chronological life-span in yeast. *FEBS Lett* 580:1903-1907
- [2] Baxter AA, Richter V, Lay FT, Poon IKH, Adda CG, Veneer PK, Phan TK, Bleackley MR, Anderson MA, Kvensakul M, Hulett MD (2015) The tomato defensin TPP3 binds phosphatidylinositol (4,5)-bisphosphate via a conserved dimeric cationic grip conformation to mediate cell lysis. *Mol Cell Biol* 35:1964-1978
- [3] Baul U, Kuroda K, Vemparala S (2014) Interaction of multiple biomimetic antimicrobial polymers with model bacterial membranes. *J Chem Phys* 141:084902

- [4] Bocchinfuso G, Palleschi A, Orioni B, Grande G, Formaggio F, Toniolo C, Park Y, Hahm K, Stella L (2009) Different mechanisms of action of antimicrobial peptides: insights from fluorescence spectroscopy experiments and molecular dynamics simulations. *J Pept Sci* 15:550-558
- [5] Bonucci A, Balducci E, Pistolesi S, Pogni R (2013) The defensin-lipid interaction: Insights on the binding states of the human antimicrobial peptide HNP-1 to model bacterial membranes. *Biochim Biophys Acta* 1828:758-764
- [6] Bulacu M, Sevink GJA (2015) Computational insight in the role of fusogenic lipopeptides at the onset of liposome fusion. *Biochim Biophys Acta* 1848: 848-858
- [7] Colavita I, Nigro E, Sarnataro D, Scudiero O, Granata V, Daniele A, Zagari A, Pessi A, Salvatore F (2015) Membrane protein 4F2/CD98 is a cell surface receptor involved in the internalization and trafficking of human β -defensin 3 in epithelial cells. *Chem Biol* 22:217-228
- [8] Cruz VL, Ramos J, Melo MN, Martinez SJ (2013) Bacteriocin AS-48 binding to model membranes and pore formation as revealed by coarse-grained simulations. *Biochim Biophys Acta* 1828: 2524-2531
- [9] de Jong DH, Singh G, Bennett WFD, Arnarez C, Wassenaar TA, Scha LV, Periole X, Tieleman DP, Marrink SJ (2013) Improved parameters for the martini coarse-grained protein force field. *J Chem Theory Comput* 9:687-697
- [10] de Medeiros LN, Domitrovic T, de Andrade PC, Faria J, Bergter EB, Weissmüller G, Kurtenbach E (2014) Psd1 binding affinity toward fungal membrane components as assessed by SPR: the role of glucosylceramide in fungal recognition and entry. *Biopolymers* 102:456-464
- [11] Efimova SS, Malev VV, Ostroumova OS (2016) Effects of dipole potential modifiers on heterogenic lipid bilayers. *J Membr Biol* 249:97-106
- [12] Ermakova EA, Faizullin DA, Idiyatullin BZ, Khairutdinov BI, Mukhamedova LN, Tarasova NB, Toporkova YY, Osipova EV, Gogolev YV, Zuev YF, Nesmelova IV (2016) Structure of scots pine defensin 1 by spectroscopic methods and computational modeling. *Int J Biological Macromol* 84: 142-152
- [13] Flinner N, Schleiff E (2015) Dynamics of the glycophorin a dimer in membranes of native-like composition uncovered by coarse-grained molecular dynamics simulations. *PLoS ONE* 10:e0133999
- [14] Gurtovenko AA, Vattulainen I (2007) Molecular mechanism for lipid flip-flops. *J Phys Chem B* 111:13554-13559
- [15] Hess B, Kutzner C, van der Spoel D, Lindahl E (2008) GROMACS 4: Algorithms for highly efficient, load-balanced, and scalable molecular simulation. *J Chem Theory Comput* 4:435-447
- [16] Hong C, Tieleman DP, Wang Y (2014) Microsecond molecular dynamics simulations of lipid mixing. *Langmuir* 30:11993-12001
- [17] Humphrey W, Dalke A, Schulten K (1996) VMD: visual molecular dynamics. *J Mol Graph* 14.1: 33-38
- [18] Janosi L, Gorfe AA (2010) Simulating POPC and POPC/POPG bilayers: conserved packing and altered surface reactivity. *J Chem Theory Comput* 6:3267-3273
- [19] Jo S, Lim JB, Klauda JB, Im W (2009) CHARMM-GUI membrane builder for mixed bilayers and its application to yeast membranes. *Biophys J* 97:50-58
- [20] Khairutdinov BI, Bessolicina EK, Efimov SV, Toporkova YY, Tarasova NB, Ermakova EA, Zuev YF (2016) Investigation of structural-dynamics properties of Scots pine defensin 1 by NMR spectroscopy. International symposium «Magnetic resonance: from fundamental research to practical applications». Book of abstracts, Kazan, p 186
- [21] Kovaleva V, Kiyamova R, Cramer R, Krynytskyy H, Gout I, Filonenko V, Gout R (2009) Purification and molecular cloning of antimicrobial peptides from Scots pine seedlings. *Peptides* 30:2136-2143
- [22] Kučerka N, Nieh MP, Katsaras J (2011) Fluid phase lipid areas and bilayer thicknesses of commonly used phosphatidylcholines as a function of temperature. *Biochim Biophys Acta* 1808: 2761-2771
- [23] Kučerka N, Holland BW, Gray CC, Tomberli B, Katsaras J (2012) Scattering density profile model of POPG bilayers as determined by molecular dynamics simulations and small-angle neutron and x-ray scattering experiments. *J Phys Chem B* 116:232-239
- [24] Kvensakul M, Lay FT, Adda CG, Veneer PK, Baxter AA, Phan TK, Poon IKH, Hulett MD (2016) Binding of phosphatidic acid by Nsd7 mediates the formation of helical defensin-lipid oligomeric assemblies and membrane permeabilization. *Proc Natl Acad Sci USA* 113:11202-11207
- [25] Lay FT, Mills GD, Poon IK, Cowieson NP, Kirby N, Baxter AA, van der Weerden NL, Dogovski C, Perugini MA, Anderson MA, Kvensakul M, Hulett MD (2012) Dimerization of plant defensin NaD1 enhances its antifungal activity. *J Biol Chem* 287:19961-19972
- [26] Lee J, Jung SW, Cho AE (2016) Molecular insights into the adsorption mechanism of human β -defensin-3 on bacterial membranes. *Langmuir* 32:1782-1790
- [27] Ludtke SJ, He K, Heller WT, Harroun TA, Yang L, Huang HW (1996) Membrane pores induced by magainin. *Biochemistry* 35:13723-13728
- [28] Marrink SJ, de Vries AH, Tieleman DP (2009) Lipids on the move: simulations of membrane pores, domains, stalks and curves. *Biochim Biophys Acta* 1788:149-168

- [29] Marrink SJ, Risselada HJ, Yefimov S, Tieleman DP, de Vries AH (2007) The MARTINI force field: coarse grained model for biomolecular simulations. *J Phys Chem B* 111:7812–7824
- [30] Midura-Nowaczek K, Markowska A (2014) Antimicrobial peptides and their analogs: searching for new potential therapeutics. *Perspect Medicin Chem* 6:73–80
- [31] Monticelli L, Kandasamy SK, Periole X, Larson RG, Tieleman DP, Marrink SJ (2008) The MARTINI coarse-grained force field: extension to proteins. *J Chem Theory Comput* 4:819–834
- [32] Periole X, Cavalli M, Marrink SJ, Ceruso MA (2009) Combining an elastic network with a coarse-grained molecular force field: structure, dynamics, and intermolecular recognition. *J Chem Theory Comput* 5:2531–2543
- [33] Poon IK, Baxter AA, Lay FT, Mills GD, Adda CG, Payne JA, Phan TK, Ryan GF, White JA, Veneer PK, van der Weerden NL, Anderson MA, Kvansakul M, Hulett MD (2014) Phosphoinositide-mediated oligomerization of a defensin induces cell lysis. *Elife* 3:e01808
- [34] Qi Y, Ingólfsson HI, Cheng X, Lee J, Marrink SJ, Im W (2015) CHARMM-GUI martini maker for coarse-grained simulations with the martini force field. *J Chem Theory Comput* 11:4486–4494
- [35] Sagaram US, El-Mounadi K, Buchko GW, Berg HR, Kaur J, Pandurangi RS, Smith TJ, Shah DM (2013) Structural and functional studies of a phosphatidic acid-binding antifungal plant defensin MtDef4: identification of an RGFRRR motif governing fungal cell entry. *PLoS ONE* 8:e82485
- [36] Sani MA, Separovic F (2016) How membrane-active peptides get into lipid membranes. *Acc Chem Res* 49:1130–1138
- [37] Sengupta D, Leontiadou H, Mark AE, Marrink SJ (2008) Toroidal pores formed by antimicrobial peptides show significant disorder. *Biochim Biophys Acta* 1778: 2308–2317
- [38] Skjerve AA, Madej BD, Dickson CJ, Teigen K, Walker RC, Gould IR (2015) All-atom lipid bilayer self-assembly with the AMBER and CHARMM lipid force fields. *Chem Commun (Camb)* 51:4402–4404
- [39] Shai Y, Oren Z (2001) From “carpet” mechanism to de novo designed diastereomeric cell-selective antimicrobial peptides. *Peptides* 22:1629–1641
- [40] Soblosky L, Ramamoorthy A, Chen Z (2015) Membrane interaction of antimicrobial peptides using E. coli lipid extract as model bacterial cell membranes and SFG spectroscopy. *Chem Phys Lipids* 187:20–33
- [41] Su Y, Li S, Hong M (2013) Cationic membrane peptides: atomic-level insight of structure–activity relationships from solid-state NMR. *Amino Acids* 44:821–833
- [42] Thevissen K, Osborn RW, Acland DP, Broekaert WF (1997) Specific, high affinity binding sites for an antifungal plant defensin on neurospora crassa hyphae and microsomal membranes. *J Biol Chem* 272:32176–32181
- [43] Thevissen K, de Mello Tavares P, Xu D, Blankenship J, Vandenbosch D, Idkowiak-Baldys J, Govaert G, Bink A, Rozental S, De Groot PW (2012) The plant defensin RsAFP2 induces cell wall stress, septin mislocalization and accumulation of ceramides in Candida albicans. *Mol Microbiol* 84:166–180
- [44] van der Spoel D, Lindahl E, Hess B, Groenhof G, Mark AE, Berendsen HJC (2005) GROMACS: fast, flexible and free. *J Comp Chem* 26:1701–1719
- [45] van der Weerden NL, Hancock REW, Anderson MA (2010) Permeabilization of fungal hyphae by the plant defensin NaD1 occurs through a cell wall-dependent process. *J Biol Chem* 285:37513–37520
- [46] van Eerden FJ, de Jong DH, de Vries AH, Wassenaar TA, Siewert J., Marrink SJ (2015) Characterization of thylakoid lipid membranes from cyanobacteria and higher plants by molecular dynamics simulations. *Biochim Biophys Acta* 1848: 1319–1330
- [47] Wang Y, Schlamadinger DE, Kim JE, McCammon JA (2012) Comparative molecular dynamics simulations of the antimicrobial peptide CM15 in model lipid bilayers. *Biochim Biophys Acta* 1818: 1402–1409
- [48] Wassenaar TA, Pluhackova K, Böckmann RA, Marrink SJ, Tieleman DP (2014) Going backward: a flexible geometric approach to reverse transformation from coarse grained to atomistic models. *J Chem Theory Comput* 10:676–690
- [49] Wu Y, He K, Ludtke SJ, Huang HW (1995) X-ray diffraction study of lipid bilayer membranes interacting with amphiphilic helical peptides: diphytanoyl phosphatidylcholine with alamethicin at low concentrations. *Biophys J* 68:2361–2369
- [50] Zhao X, Yu H, Yang L, Li Q, Huang X (2015) Simulating the antimicrobial mechanism of human β -defensin-3 with coarse-grained molecular dynamics. *J Biomol Struct Dyn* 33:2522–2531